

## Molecular Approach Towards Screening of Biological Targets of Berberine and its Production Sources

Jhansi Preetham Garrepelly<sup>1</sup>, Kiranmayi P Y N L<sup>1</sup>, Thanuja Bandari<sup>1</sup> and Rajeswara Reddy Erva<sup>1\*</sup>

<sup>1</sup>Department of Biotechnology, National Institute of Technology Andhra Pradesh, Tadepalligudem, West Godavari (D), Andhra Pradesh-534101, India.

\*Corresponding Author : erreddy@nitandhra.ap.in

### Abstracts

Traditional medicine dating back to centuries ago has been used to treat several ailments of humans. Berberine is a chemical compound that is found in plants which is being used in the form of drugs to work against various diseases. For optimum production of Berberine suitable plant species must be selected out of *Coptis japonica* and *Argemone mexicana* which are high yielding. From the pathway of Berberine production it is known that more the amount of substrate Canadine gives more amount of Berberine. The objective of this work is to compare the Canadine production efficiency among *Coptis japonica* and *Argemone mexicana* by molecular level studies for both the species. It is concluded that the enzyme, (S)-canadine synthase from *Argemone mexicana* is more efficient towards the production of (S)-canadine which in turn lead to Berberine production. Later the efficiency of Berberine against various disease targets is tested by molecular docking approach under same experimental conditions. Molecular docking results conferred the efficiency of Berberine in the treatment of gastric cancer over the other diseases.

**Keywords:** Berberine, Canadine, Canadine Synthase, Homology Modeling and Molecular Docking.

### Introduction

Plants have been used for medicinal purposes long before prehistoric period. Chemical compounds extracted from plants have been

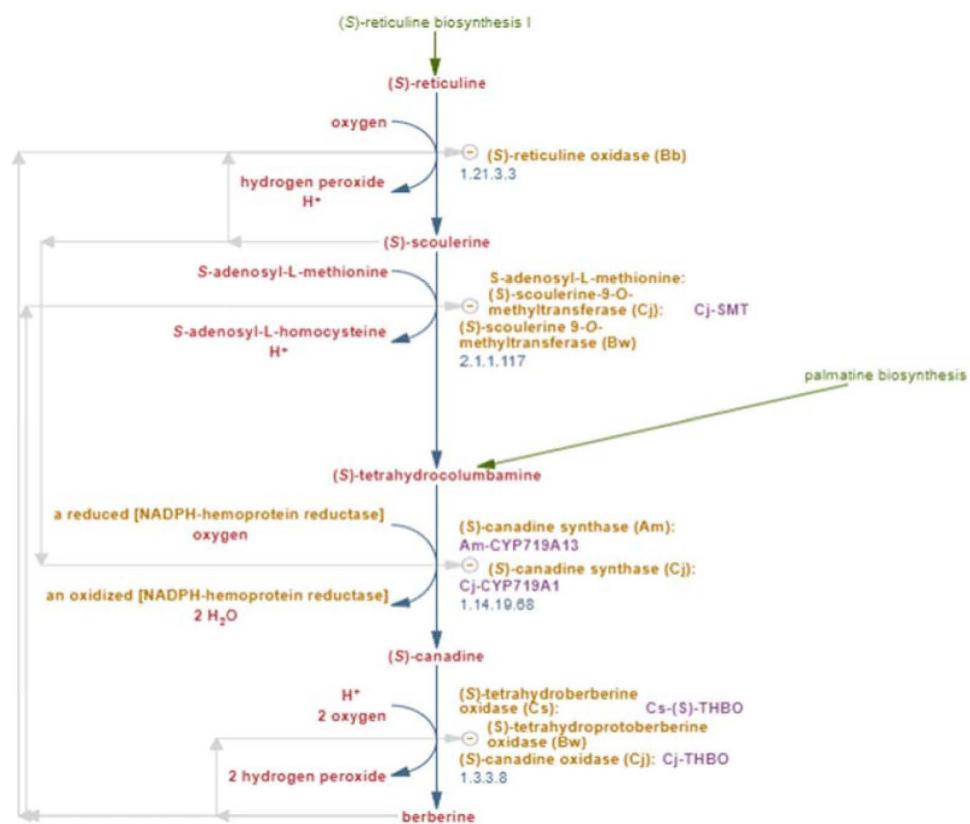
known to counteract a number of ailments (1). Even though herbal medicines have lesser potent when compared to synthetic drugs in some cases, still they are considered lesser toxic or having lesser side effects in contrast to synthetic drugs. According to statistical data provided by World Health Organization (WHO), 80 percent of people worldwide rely on herbal medicines for some aspect of their primary health care needs. WHO also estimated that around 21,000 plant species have the potential for being used as medicinal plants for a wide range of diseases (2). Fast developing countries such as India and China are contributing as much as 80% of total natural sources for manufacturing of drugs, which are being used worldwide. According to the World Bank reports, trade in medicinal plants, botanical drug products and raw materials are growing at an impressive annual growth rate between 5 and 15%. In India the value of botanicals related trade is about US \$10 billion per annum while the annual export of US is \$1.1 billion. China's annual herbal drug production is worth a huge amount of US\$48 billion with export of US \$3.6 billion (3).

The observation is that drugs got from plants handled in unrefined structure without the segregation of active molecules would be safer (4). As the ultimate aim for any medicine is their non-toxicity, effectiveness, stability, specificity, and potency, herbal medicines are highly encouraged (5). Consistency in biological activity and composition are essential requirements for the safe and effective use of therapeutic drugs (3).

Berberine is one of those herbal medicines found in several plants (6). Berberine (2, 3-methylenedioxy-9, 10-dimethoxy-protoberberine) is a chemical found in several plants which is a cure for a wide range of diseases. Berberine is a quaternary ammonium salt from the protoberberine group of benzylisoquinoline alkaloids (5). Benzylisoquinoline alkaloids (BIAs) are a large, diverse group of ~2500 specialized plant metabolites (2). Among them, the natural isoquinoline alkaloid Berberine has been employed in Ayurvedic and Chinese Medicine for hundreds of years with a wide range of pharmacological and biochemical effects (8). Berberine has a wide range of beneficial properties including anti-cancer, anti-inflammatory, anti-bacterial and cholesterol lowering effects.

Berberine has shown impressive efficacy against various bacterial strains like cholera, giardia, shigella, and salmonella. It has the potential to work against even staphylococcus, streptococcus, and clostridium (9). Some of the most common side effects of Berberine usage include vomiting, hypertension (high blood pressure), nausea, chest pain, skin rashes, breathing problems and skin hives (10).

Berberine has been isolated from various plant families including Papaveraceae, Berberidaceae, Fumariaceae, Menispermaceae, Ranunculaceae, Rutaceae and Annonaceae (11). Berberine is found in significant amounts in *Coptidis rhizome*, *Coptis japonica*, *Coptis chinensis*, *Argemone Mexicana*, *Berberis aristata*,



**Fig. 1 : Berberine Biosynthesis Pathway**  
(Image courtesy: MetaCyc Pathway - Berberine biosynthesis)

*Berberis aquifolium*, *Berberis asiatica*, *Berberis croatica*, *Berberis thunbergii* and *Berberis vulgaris* (9). Out of all the species producing Berberine, *Coptis japonica* and *Argemone mexicana* are the two species having considerable amount in them (12,13).

**(S)-Canadine Synthase:** Canadine is the last but one product in the pathway of production of Berberine (Figure 1). (S)-canadine synthase (EC 1.14.21.5) is an enzyme that catalyzes the chemical reaction:

**(S)-Tetrahydrocolumbamine + NADPH + H<sup>+</sup> + O<sub>2</sub> →(S)-canadine+ NADP<sup>+</sup> + 2 H<sub>2</sub>O:** Concludes

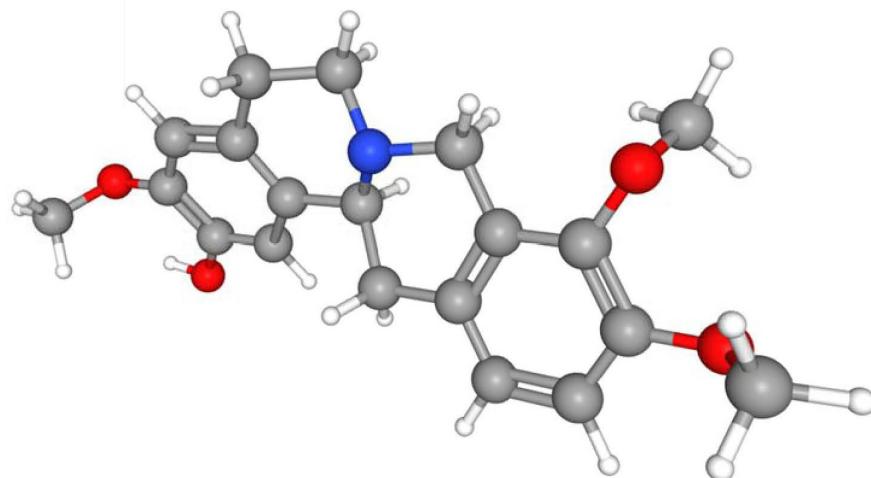
that increase in the amount of Canadine leads to increase in Berberine production confirming Canadine as a non-substrate inhibitor (8). Hence this study is aimed towards the prediction of Canadine production efficiency from the two high yielding Berberine plant producers namely, *Coptis japonica* and *Argemone mexicana*.

#### Materials and Methods

**Sequence and Structure Retrieval:** Amino acid sequences of (S)-canadine synthase from the species *Coptis japonica* (Accession No: Q948Y1) and *Argemone Mexicana* (Accession No: B1NF19) are retrieved from NCBI in FASTA format. The 3D structure of Tetrahydrocolumbamine is taken from

**Table 1:** Targets of seven chosen diseases

S.No	Receptor	Receptor's PDB Code	Clinical Applications
1	Epidermal growthfactor receptor	1JL9	Breast cancer
2	Interleukin 6	1P9M	Skin aging
3	Wee1 like protein kinase	1X8B	Gastric cancer
4	BCL2 associated X protein	4ZGM	Gastric cancer
5	Acetyl-Co A carboxylase	4ASI	Hypercholesterolemia
6	ABCG2 Transporter	5DO7	Breast cancer



**Fig. 2 :** 3D structure of Tetrahydrocolumbamine

PubChem database with the confirm ID CID\_440229 (figure 2). All the disease targets or receptors of various diseases like Breast cancer (1JL9, 5DO7), Skin aging (1P9M), Gastric cancer (1X8B, 4ZGM) and Hypercholesterolemia (4ASI) where the Berberine has proved its efficacy are retrieved from PDB (Table 1) (14).

**Sequence Alignment and Structural Analysis of *Coptis japonica* and *Argemona Mexicana*:** Local and global alignments between both the sequences is performed to find the sequence similarity pattern existing between them (15). This is followed by the primary and secondary structural analysis of (S)-canadine synthase from *Coptis japonica* and *Argemona Mexicana* to predict the physicochemical properties and the nature of secondary structural composition of the enzyme in both the plant species using ProtParam (16), SOPMA (17) and GOR4 (18) tools.

**Structural Modelling and Validation:** BLAST and PSI-BLAST were performed against PDB database for (S)-canadine synthase from *Coptis japonica* and *Argemona Mexicana* to find out the presence of any similar type of proteins whose structures are well studied.

Modelling of enzyme tertiary structures is done using I-TASSER [19] and PHYRE2[20] tools. The modelled enzyme structures from I-TASSER and PHYRE2 were validated by ProQ (21), PROCHECK (22), Verify3D [23], DFIRE Energy (24) and ERRAT (25) tools.

**Energy Minimization and Molecular Docking:** To proceed with the screening of best plant producer for Canadine which in turn gives Berberine among *Coptis japonica* and *Argemona Mexicana* based on molecular docking approach, the energy minimization of the modelled proteins and the substrate Tetrahydrocolumbamine is done using YASARA online web server tool (26). Berberine potency against various disease targets was studied by molecular docking approach using various tools like FRED (27), GOLD (28), Patch Dock (29), Surflex-Dock (30,31), AutoDock-Vina (32), Libdock and CDOCKER (33) etc., Unlike the

above cases of docking performed in different experimental conditions using different tools where in the comparison of Berberine efficiency against the disease target becomes difficult, a total of six disease targets were allowed to interact with Berberine compound in an unified environment to easily analyze the potency of compound based on the dock score obtained. Energy minimization of the various disease receptors is done using UCSF chimera tool [34]. All molecular docking runs are executed in a flexible docking wizard called iGEMDOCK (35).

## Results and Discussion

**Structural Analysis of (S)-canadine synthase:** Primary structural analysis of (S)-canadine synthase from *Coptis japonica* and *Argemona Mexicana* by PROTPARAM tool (Table 2) has confirmed both enzymes as positively charged, thermo stable and non-hydrophobic in nature based on the number of positive residues, aliphatic index and the GRAVY<sup>a</sup> scores they possess. However, (S)-canadine synthase from *Coptis japonica* was stable and the same enzyme from *Argemona Mexicana* was unstable based on their instability index scores.

Secondary structure analysis of (S)-canadine synthase from *Argemona mexicana* and *Coptis japonica* is performed using GOR4 and SOPMA tools (Table 3). This analysis revealed that, both the enzymes were rich in Alpha Helices and Random coils. They have a little Extended strand portion as their secondary structural element and the amount of Beta turns they possess was very little.

**Sequence Alignment:** In order to predict the similarity levels between the (S)-canadine synthase from *Coptis japonica* and *Argemona Mexicana* local and global sequence alignments were performed. This resulted into an identity score of 65.4% and 63.3% respectively (less than 70%). Therefore, both the amino acid sequences are sufficiently distinct for a chance of possessing different functionalities as enzymes.

**Structural Modelling and Validation:** As no experimental structures for (S)-canadine synthase

**Table 2:** PROTPARAM results for *Coptis japonica* and *Argemona mexicana*

PARAMETER	ORGANISM	
	<i>Coptis japonica</i>	<i>Argemona mexicana</i>
Number of amino acids	491	504
Molecular weight (g/mole)	55352.51	57452.49
Theoretical isoelectric point(pi)	8.69	9.14
Total number of negatively charged residues (Asp+Glu)	52	50
Total number of positively charged residues (Arg+Lys)	57	59
Instability index	37.06	44.74
Aliphatic index	96.17	93.23
GRAVY <sup>a</sup>	-0.053	-0.217

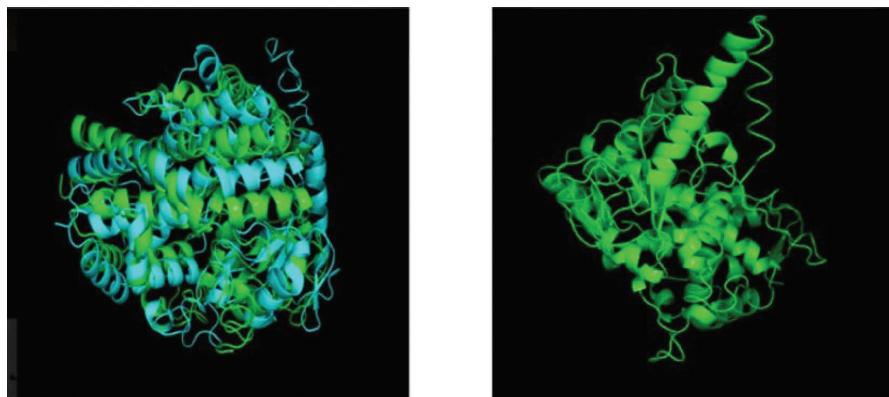
**Table 3:** GOR4 and SOPMA results for *Coptis japonica* and *Argemona Mexicana*

ORGANISM	SECONDARY STRUCTURE ANALYSIS TOOLS		
	PARAMETERS	GOR 4	SOPMA
<i>Argemona mexicana</i>	Alpha Helix	43.63%	45.63%
	Beta turn	0	4.17%
<i>Coptis japonica</i>	Extended strand	11.51%	11.71%
	Random coil	42.86%	38.49%
	Alpha helix	47.66%	47.66%
	Beta turn	0%	4.07%
	Extended strand	11.41%	9.98%
	Random coil	40.94%	38.29%

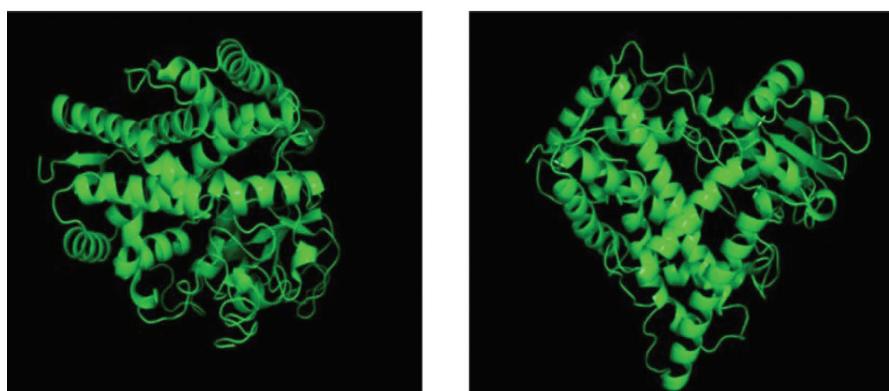
are available in biological databases till date, the enzyme's structure should be modelled by appropriate modelling tool. To choose the with the suitable modelling method, initially BLASTP search was performed to find the suitable templates with high level of sequence similarity. As PSI-BLAST is more sensitive than BLAST in finding distantly related sequences that are missed in a BLAST search, PSI-BLAST has also been performed to obtain accurate hits (36).

The percentage identity values of the first five hits obtained from BLASTp and PSI-BLAST

of both the amino acid sequences of the (S)-canadine synthase from the species *Argemona mexicana* and *Coptis japonica* are all less than 30. As it resulted into very less percentage of sequence identity, where homology modelling approach is not suitable to proceed further, ab initio method is opted for tertiary structure modeling of the enzyme (S)-canadine synthase from Argemona Mexicana (Figure 3) and Coptis japonica (Figure 4) using I-TASSER and PHYRE2 tools.



**Fig. 3:** Modeled structures of (S)-canadine synthase from *Argemona mexicana* using I-TASSER (a) and PHYRE2 (b) tools



**Fig. 4:** Modeled structures of (S)-canadine synthase from *Coptis japonica* using I-TASSER (a) and PHYRE2 (b) tools

The modeled structures of the enzyme (S)-canadine synthase from *Argemona mexicana* and *Coptis japonica* from both I-TASSER and PHYRE2 are validated using tools like ERRAT, ProQ, Verify 3D, DFIRE Energy (<http://www.scbio-iitd.res.in/software/proteomics/protsav.jsp>) (Table 4).

Based on results of all the above validation tools I-TASSER models were concluded as the best models for enzyme (S)-canadine synthase from *Argemona mexicana* and *Coptis japonica*. Hence, the modelled enzyme structures from I-TASSER tool were revalidated through

Ramachandran plot analysis using Procheck (Table 5) tool. Having more than 97% of the residues in the favored and allowed regions both the I-TASSER modelled structures proved their reliability for the further use.

**Molecular Docking:** Stable and flexible docking has been performed using iGEMDOCK between the energy minimized structures of Tetrahydrocolumbamine and ab initio modelled (S)-canadine synthase from *Argemona mexicana* and *Coptis japonica* as ligand and receptors respectively (table 6).

**Table 4 :** Comparison of PHYRE2 and I-TASSER structures

Model	<i>Coptis japonica</i>		<i>Argemona mexicana</i>		
Validation	PHYRE 2	I-TASSER	PHYRE 2	I-TASSER	Remarks
<b>Tools</b>					CORRECT : LG score > 1.5 Max Sub > 0.1
<b>ProQ</b>	LG score: 4.548 Max Sub: 0.103	LG score: 6.263 Max Sub: 0.209	LG score: 4.005 Max Sub: 0.117	LG score: 6.162 Max Sub: 0.211	GOOD: LG score > 3 Max Sub > 0.5
<b>Verify 3D</b>	132.5601	156.0701	120.09	178.6701	VERY GOOD: LG score > 5 Max Sub > 0.8
<b>DFire Energy</b>	-978.98	-1116.54	-1087.27	-1171.88	Higher the quality score, higher the quality
<b>ERRAT</b>	51.1161	79.7101	68.145	86.492	Lower the energy, better the quality of structure Good resolution- 95% or higher lower resolutions- around 91%

**Table 5:** Ramachandran plot parameters for I-TASSER structures

Procheck	<i>Coptis japonica</i>	<i>Argemona mexicana</i>
<b>Number of residues in favored region</b>	72.7%	72.1%
<b>Number of residues in allowed region</b>	25%	25.2%
<b>Number of residues in outlier region</b>	2.3%	2.8%

**Table 6 :** Fitness values from iGEMDOCK

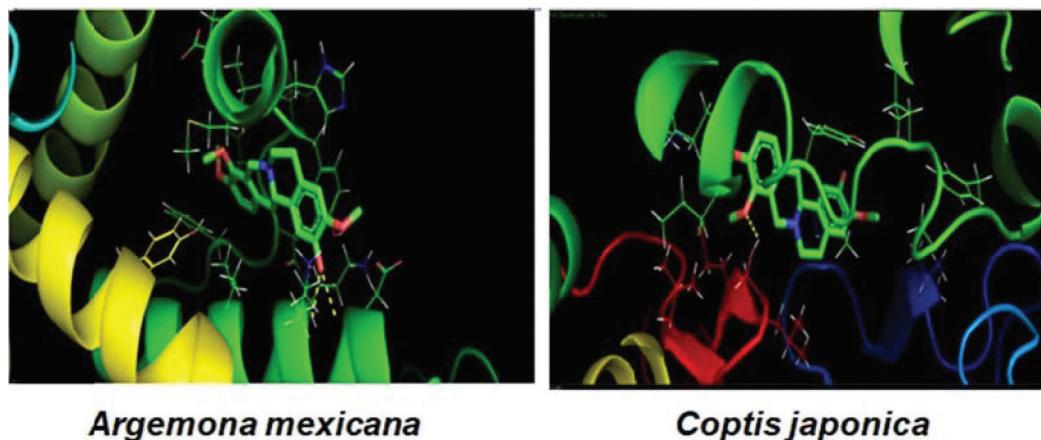
LIGAND	RECEPTOR ((S)-canadine synthase)	Docking Score
Tetrahydrocoloumbamine	<i>Argemona mexicana</i> <i>Coptis japonica</i>	-102 kcal/mole -98.9431 kcal/mole

The best docked poses are obtained between the modeled enzymes along with respective fitness values (Figure 5). Lower the free energy better is the binding between the ligand and receptor. As the free energy of *Argemona mexicana*'s (S)-canadine synthase against Tetrahy- drocoloumbamine is lower than that of *Coptis japonica*, it can be concluded that the enzyme, (S)-canadine synthase from *Argemona mexicana* is more efficient.

Further proceeding with determining the Berberine efficiency as a therapeutic agent against various diseases (37-45), stable docking is performed for all the six disease targets individually

against Berberine as ligand, under same experimental conditions (Figure 6, Table 7).

From the above tabulated values, the docking energy for 1X8B (-115.301 kcal/mole) is the least. 1X8B is the disease target for gastric cancer. Based on the docking results, out of all the six disease targets, the Wee1 like protein kinase is efficiently blocked by Berberine. The inhibition of Wee1 like protein kinase leads to premature mitosis and cell death. Berberine may act as an effective therapeutic drug for the treatment of gastric cancer. Therefore, Berberine is most efficient in acting against this disease target when compared to others.



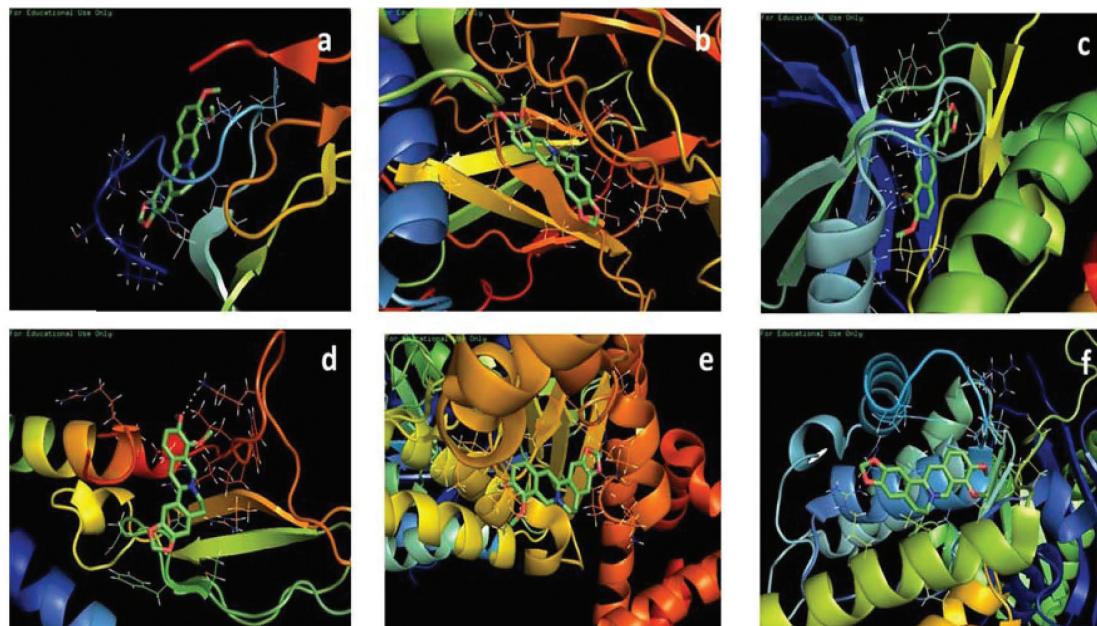
**Fig. 5:** Interaction of Tetrahydro-columbamidine with modeled structures

**Table 7:** Docking energy values for six receptors against Berberine

S.No	Receptor	iGEMDOCK Docking energy (kcal/mole)
1	1JL9	-83.05
2	1P9M	-105.48
3	1X8B	-115.301
4	4ZGM	-89.3
5	4ASI	-102.35
6	5DO7	-94.3

### Conclusion

*Coptis japonica* and *Argemona mexicana* are the two Berberine high yielding plant species. (S) canadine synthase is an enzyme that catalyzes the reaction of Canadine production from which Berberine is synthesized. Global and local alignments between the enzyme sequences from both the species confirmed that they are distant from each other. The structural analysis confirmed that both the species are producing the same kind of enzyme with most of the similar characteristics. Molecular docking of the validated and modelled



**Fig. 6 :** Best docked pose of Berberine against 1JL9 (a), 1P9M (b), 1X8B (c), 4ZGM (d), 4ASI (e) and 5DO7 (f)

structures with Tetrahydrocolumbamine has proved *Argemone mexicana* as a potent Canadine producer over *Coptis japonica* which in turn produce Berberine, with the energy value of -102 kcal/mole. The subsequent docking runs of Berberine against its disease targets confirmed the higher therapeutic capacity of Berberine in treatment of Gastric Cancer over the breast cancer, skin ageing and hypercholesterolemia.

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**Conflict of interest:** Authors have no conflict of interest regarding the publication of paper.

## References

1. Tillhon M, Ortiz LMG, Lombardi P, Scovassi AI. (2012). Berberine: new perspectives for old remedies. Biochemical pharmacology, 84: 1260-1267.
2. [www.nhp.gov.in/introduction-and-importance-of-medicinal-plants-and-herbs\\_mtl](http://www.nhp.gov.in/introduction-and-importance-of-medicinal-plants-and-herbs_mtl)
3. Patwardhan B, Warude D, Pushpangadan P, Bhatt N. (2005). Ayurveda and traditional Chinese medicine: a comparative overview. Evidence-Based Complementary and Alternative Medicine, 2(4):465-473.
4. Manohar PR. (2014). Toxicity of Ayurveda medicines and safety concerns: The need to revive the branch of toxicology in Ayurveda. Ancient science of life, 34(1): 1-2.

5. SL NBSAR. (2017). Comparison of medicinally important natural products versus synthetic drugs-a short commentary. *Nat Prod Chem Res*, 6(308): 2.
6. Tang J, Feng Y, Tsao S, Wang N, Curtain R, Wang Y. (2009). Berberine and Coptidis rhizoma as novel antineoplastic agents: a review of traditional use and biomedical investigations. *Journal of ethnopharmacology*, 126: 5-17.
7. Nechepurenko I, Salakhutdinov N, Tolstikov G. (2010). Berberine: Chemistry and biological activity. *Chemistry for Sustainable Development*, 18: 1-23.
8. Branton A, Jana S. (2017). The use of novel and unique biofield energy healing treatment for the improvement of poorly bioavailable compound, berberine in male Sprague Dawley rats. *American Journal of Clinical and Experimental Medicine*, 5(4).
9. Vuddanda PR, Chakraborty S, Singh S. (2010). Berberine: a potential phytochemical with multispectrum therapeutic activities. *Expert opinion on investigational drugs*, 19: 1297-1307.
10. <https://www.webmd.com/vitamins/ai/ingredientmono-1126/berberine>
11. Grycova L, Dostál J, Marek R. (2007). Quaternary protoberberine alkaloids. *Phytochemistry*, 68: 150-175.
12. Sato F, Yamada Y. (1984). High berberine-producing cultures of *Coptis japonica* cells. *Phytochemistry*, 23: 281-285.
13. Singh S, Singh TD, Singh VP, Pandey VB. (2010). Quaternary alkaloids of *Argemone mexicana*. *Pharmaceutical biology*, 48: 158-160.
14. Bank PD. (1971). Protein data bank. *Nature New Biol*, 233: 223.
15. Walker JM. (2005). *The proteomics protocols handbook*. Springer.
16. Geourjon C, Deleage G. (1995). SOPMA: significant improvements in protein secondary structure prediction by consensus prediction from multiple alignments. *Bioinformatics*, 11: 681-684.
17. Kouza M, Faraggi E, Kolinski A, Kloczkowski A. (2017). The GOR method of protein secondary structure prediction and its application as a protein aggregation prediction tool. *Prediction of Protein Secondary Structure*: Springer, 7-24.
18. Yang J, Zhang Y. (2015). Protein structure and function prediction using I-TASSER. *Current protocols in bioinformatics*, 52: 1-15.
19. Kelley LA, Mezulis S, Yates CM, Wass MN, Sternberg MJ. (2015). The Phyre2 web portal for protein modeling, prediction and analysis. *Nature protocols*, 10: 845-858.
20. Colovos C, Yeates TO. (1993). Verification of protein structures: patterns of nonbonded atomic interactions. *Protein science*, 2: 1511-1519.
21. Uziela K, Menendez Hurtado D, Shu N, Wallner B, Elofsson A. (2017). ProQ3D: improved model quality assessments using deep learning. *Bioinformatics*, 33: 1578-1580.
22. Lüthy R, Bowie JU, Eisenberg D. (1992). Assessment of protein models with three-dimensional profiles. *Nature*, 356: 83-85.
23. Yang Y, Zhou Y. (2008). Specific interactions for ab initio folding of protein terminal regions with secondary structures. *Proteins: Structure, Function, and Bioinformatics*, 72: 793-803.
24. Xu B, Yang Y, Liang H, Zhou Y. (2009). An all atom knowledge based energy function for protein-DNA threading, docking decoy discrimination, and prediction of transcription factor binding profiles. *Proteins: Structure, Function, and Bioinformatics*, 76: 718-730.

25. Hsu K-C, Chen Y-F, Lin S-R, Yang J-M. (2011). iGEMDOCK: a graphical environment of enhancing GEMDOCK using pharmacological interactions and post-screening analysis. *BMC bioinformatics*, 12: S33.
26. Krieger E, Vriend G. (2014). YASARA View-molecular graphics for all devices-from smartphones to workstations. *Bioinformatics*, 30: 2981-2982.
27. Al-masri IM, Mohammad MK, Tahaa MO. (2009). Inhibition of dipeptidyl peptidase IV (DPP IV) is one of the mechanisms explaining the hypoglycemic effect of berberine. *Journal of enzyme inhibition and medicinal chemistry*, 24: 1061-1066.
28. Liu B, Fu X-Q, Li T, et al. (2017). Computational and experimental prediction of molecules involved in the anti-melanoma action of berberine. *Journal of ethnopharmacology*, 208: 225-235.
29. Saha SK, Khuda-Bukhsh AR. (2014). Berberine alters epigenetic modifications, disrupts microtubule network, and modulates HPV-18 E6–E7 oncoproteins by targeting p53 in cervical cancer cell HeLa: a mechanistic study including molecular docking. *European Journal of Pharmacology*, 744: 132-146.
30. Shan Y-Q, Ren G, Wang Y-X, et al. (2013). Berberine analogue IMB-Y53 improves glucose-lowering efficacy by averting cellular efflux especially P-glycoprotein efflux. *Metabolism*, 62: 446-456.
31. Ji H-F, Shen L. (2012). Molecular basis of inhibitory activities of berberine against pathogenic enzymes in Alzheimer's disease. *The Scientific World Journal*, 1-4.
32. Yasmeen S. (2017). Exploring thermodynamic parameters and the binding energetic of berberine chloride to bovine serum albumin (BSA): Spectroscopy, isothermal titration calorimetry and molecular docking techniques. *Thermochimica Acta*, 655: 76-86.
33. Ling Y, Ling-Ling W, Qian L, et al. (2018). Novel berberine derivatives: Design, synthesis, antimicrobial effects, and molecular docking studies. *Chinese journal of natural medicines*, 16: 774-781.
34. Pettersen EF, Goddard TD, Huang CC, et al. (2004). UCSF Chimera—a visualization system for exploratory research and analysis. *Journal of computational chemistry*, 25: 1605-1612.
35. Reddy ER, Babu RS, Chandrasai PD, Madhuri P. (2016). Exploration of the binding modes of L-asparaginase complexed with its amino acid substrates by molecular docking, dynamics and simulation. *3 Biotech*, 6: 105.
36. Galanis S, Smolke CD. (2015). Optimization of yeast-based production of medicinal protoberberine alkaloids. *Microbial cell factories*, 14: 1-13.
37. Zhao Y, Ma J, Fan Y, et al. (2018). TGF  $\alpha$  transactivates EGFR and facilitates breast cancer migration and invasion through canonical Smad3 and ERK/Sp1 signaling pathways. *Molecular oncology*, 12: 305-321.
38. Fukuda K, Hibiya Y, Mutoh M, Koshiji M, Akao S, (1999). Fujiwara H. Inhibition of activator protein 1 activity by berberine in human hepatoma cells. *Planta Medica*, 65: 381-383.
39. Kim S, Kim Y, Kim JE, Cho KH, Chung JH. (2008). Berberine inhibits TPA-induced MMP-9 and IL-6 expression in normal human keratinocytes. *Phytomedicine*, 15: 340-347.
40. Fagot D, Asselineau D, Bernerd F. (2002). Direct role of human dermal fibroblasts and indirect participation of epidermal keratinocytes in MMP-1 production after UV-B irradiation. *Archives of dermatological research*, 293: 576-583.

41. Beetz RP, T. Oppel, W. Kaffenberger, RA Rupec, M. Meyer, D. Van Beuningen, P. Kind, G. Messer, A. (2000). NF- $\kappa$ B and AP-1 are responsible for inducibility of the IL-6 promoter by ionizing radiation in HeLa cells. International journal of radiation biology, 76: 1443-1453.
42. Squire CJ, Dickson JM, Ivanovic I, Baker EN. (2005). Structure and Inhibition of the Human Cell Cycle Checkpoint Kinase, Wee1A Kinase: An Atypical Tyrosine Kinase with a Key Role in CDK1 Regulation. Structure, 13: 541-550.
43. Li J, Yanyan M, Mu L, Chen X, Zheng W. (2019). The expression of Bcl-2 in adenomyosis and its effect on proliferation, migration, and apoptosis of endometrial stromal cells. Pathology-Research and Practice, 215: 152477.
44. Kim JB, Ko E, Han W, Shin I, Park SY, Noh D-Y. (2008). Berberine diminishes the side population and ABCG2 transporter expression in MCF-7 breast cancer cells. Planta Medica, 74: 1693-1700.
45. Nakanishi T, Ross DD. (2012). Breast cancer resistance protein (BCRP/AGCB2): Its role in multidrug resistance and regulation of its gene expression. Chinese Journal of Cancer, 31(2): 73-99.