

DNA Barcoding and Phylogenetic Analysis of Indian Medicinal Plant *Piper longum*

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Abstract

Piper longum is an important medicinal plant, widely used in treating diseases in the Ayurvedic system. *Piper longum* is dioecious with male and female plants. Female *Piper longum* is more widely used than the male owing to its metabolites, making it economically valuable. This study aimed to amplify the DNA barcodes for both male and female plants using *rbcL*, ITS2, and *psbA-trnH* followed by Sanger sequencing. The phylogenetic trees were constructed using the UPGMA method from the sequence data obtained. Based on the sequence alignment, *rbcL* and *psbA-trnH* were found to be effective in distinguishing *Piper longum* between or within the species when compared to ITS2. While complete *rbcL* and *psbA-trnH* regions were amplified, ITS2 showed incomplete amplification for *Piper longum*. Hence, *rbcL* and *psbA-trnH* can be used for the successful authentication of *Piper longum*.

Keywords: *Piper longum*, *rbcL*, ITS2, *psbA-trnH*, Sanger sequencing, Phylogenetic trees

Introduction

Piper longum (Piperaceae family) commonly called “long pepper” or “Pippali” is a dioecious medicinal plant species that is widely distributed in tropical and subtropical regions of

most Asian countries like India, Nepal, Sri Lanka, and Indonesia. It is very difficult to determine the sex of the plant at the earlier stage since the male and female *Piper longum* plants are morphologically very similar until the formation of spikes. Male spikes are long greenish-yellow coloured, fleshy, cylindrical, with minute flowers, and female spikes are short yellow-coloured, erect, thick, and shorter than male spikes (1). Studies have proved that the female spikes of *Piper longum* contain more valuable compounds than male spikes (2), hence female plants are preferred over male plants. Thus, the elimination of unwanted plants is necessary (3). Piperine is an important alkaloid present in the fruits of *Piper longum* which contains high medicinal properties. It also contains piperidine, piperlongumine, lignans, volatile oils, esters, organic acids, and minerals such as calcium, phosphorus, and iron (4).

The parts of the *Piper longum* plant such as spikes, roots, and stems are commonly used for pharmacological purposes. The metabolites present in the roots, spikes, and leaves of the *Piper longum* plant are used for several diseases, especially for respiratory-related problems and also for worms, dyspepsia, and amoebiasis (5).

Nowadays, traditional crops are replaced by medicinal and aromatic plants. India plays a vital role in manufacturing and exporting herbal product-based medicines globally. Mainly, the fruits and roots of *P. longum* are exported in fresh, dried, or powdered form which is much more expensive to many countries such as the USA, Canada, Philippines, Europe, Australia, and Southeast Asian countries like Singapore, Japan, Pakistan, etc. Hence the authenticity of these medicinal plants is a matter of importance (6).

Many techniques have been evolved to identify the diversity in dioecious medicinal plant species. Molecular markers such as Random Amplified Polymorphic DNA (RAPD) (7), Inter Simple Sequence Repeats (ISSR) (8), and DNA barcoding are widely used. Among them, DNA barcoding (9) is one of the most reliable methods for the identification of different types of *Piper* species. It uses universal primers (DNA barcodes) to amplify highly conserved regions of DNA fragments of either nuclear (ITS) or organellar (chloroplast genes such as *rbcl* and *psbA-trnH*) genomes. Compared to other traditional methods it provides more accurate results (10).

Materials and Methods

DNA isolation and quantification

The male and female *Piper longum* (long pepper) plants were collected from Aromatic and Medicinal Plants Research, Station Odakkali, Ernakulam district, Kerala. The young leaves of male and female *Piper longum* plants were collected and washed properly under running tap water and air-dried. The male and female plant leaves were weighed and ground using liquid nitrogen to a powder. To this, CTAB extraction buffer and β -mercaptoethanol were added and incubated at 60°C for 30 minutes. This was then centrifuged at 10,000 rpm for 10 minutes. The supernatant was transferred into new centrifuge tubes and treated with RNase A and incubated at 32°C for 20 minutes. To this, 2/3 volume of chloroform: isoamyl alcohol (24:1) was added

and centrifuged at 10,000 rpm for 10 minutes. The above step was repeated twice. The aqueous phase was transferred to new tubes and DNA was precipitated using isopropanol. The precipitated DNA was washed with 70% ice-cold ethanol by centrifuge at 10,000 rpm for another 5 minutes. The DNA pellet was then air-dried and resuspended in TE buffer and stored in a deep freezer at -4°C (11). The concentration and purity of the DNA samples were determined in NanoDrop.

DNA barcoding

DNA amplification was carried out using three different primers *rbcl*, ITS2, and *psbA-trnH*. The reaction mixture for PCR amplification included genomic DNA (50 ng), forward and reverse primers each (10 pmol), master mix, and nuclease-free water. The conditions for *rbcl* amplification are as follows; initial denaturation at 94°C for 4 minutes followed by 35 cycles of denaturation at 94°C for 30 seconds, annealing temperature at 55°C for 30 seconds, extension at 72°C for a minute and final extension 72°C for 10 minutes. For ITS2, the amplification was set as initial denaturation at 95°C for 3 minutes followed by 35 cycles of denaturation at 92°C for a minute, annealing temperature at 50°C for a minute, extension at 72°C for 2 minutes, and a final extension at 72°C for 10 minutes. Amplification of *psbA-trnH* was performed under the following conditions; initial denaturation at 95°C for 3 minutes followed by 35 cycles of denaturation at 95°C for 30 seconds annealing temperature at 55°C for a minute, extension at 72°C for 1.5 minutes and a final extension at 72°C for 4 minutes. The amplified products were visualized by 1.2% Agarose gel electrophoresis in 1x Tris-EDTA buffer and documented using a gel documentation system (Alpha Innotech, USA). The PCR amplified products were purified and sequenced using Sanger sequencing (Agrigenome labs, Kochi, Kerala).

Sequence analysis

The amplified sequences were analyzed using the NCBI- BLAST. The phylogenetic trees

were constructed using the UPGMA method and the Maximum Composite Likelihood model for 1000 bootstraps using MEGA-X software with the most similar *Piper species* in BLAST.

Results and Discussion

Quantification of DNA

The DNA concentration and its purity are important in molecular studies for obtaining accurate results. The concentration of DNA was found to be between 500.1 ng/ μL and 144.1 ng/ μL . The purity (260/280 nm) was found to be in the range of 1.89 and 1.86. The DNA samples of good quality which ranged between 1.8-1.9 were considered for PCR amplification.

DNA barcoding

The size of the amplified product for *rbcL* was ~600 bps for both male and female plants (Figure 1).

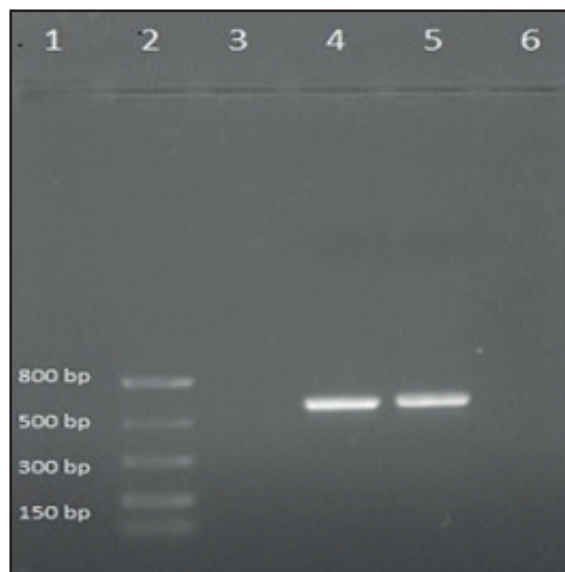


Figure 1: *rbcL* region amplification of *Piper longum*; Lane 2 is the DNA ladder, Lane 4 is the male plant, and Lane 5 is the female plant.

For ITS2, a clear band was seen in the case of the male sample of about 300-400 bps. ITS2 amplification in the female sample was unsuccessful under the given conditions (Figure 2).

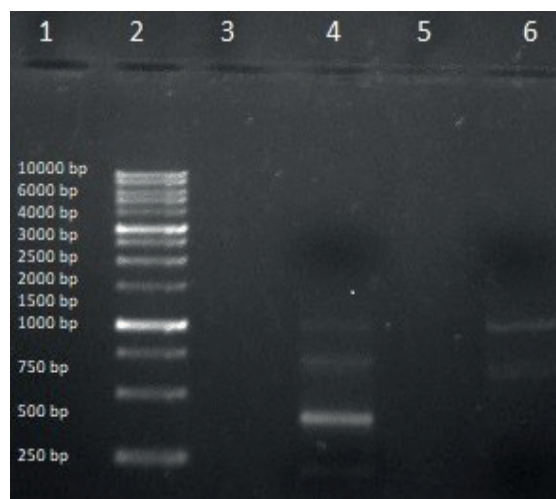


Figure 2: ITS2 region amplification of *Piper longum*; Lane 2 is the DNA ladder, Lane 4 is the male plant, and Lane 6 is the female plant.

For *psbA-trnH*, a clear intact band in both male and female samples showed the presence of DNA which was of band size ~500 bps (Figure 3).

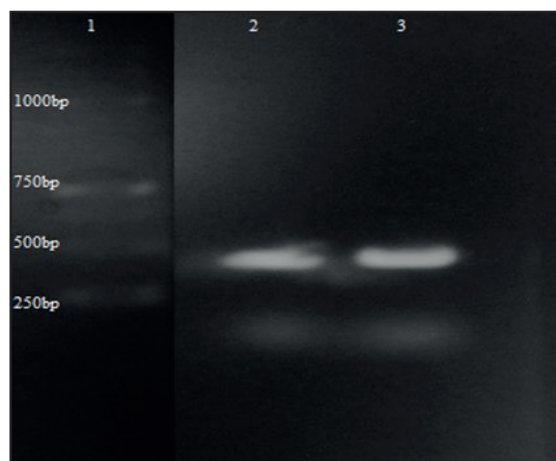


Figure 3: *psbA-trnH* region amplification of *Piper longum*; Lane 1 is the DNA ladder, Lane 2 is the male plant, and Lane 3 is the female plant.

Both *rbcL* and *psbA-trnH* regions have shown clear and consistent bands for both male and female plants. However, in the case of ITS2, amplification was successful only in the

male sample. Studies have shown that the ITS region shows more variation than chloroplast DNA markers. The polymorphic behaviour of ITS could be due to either introgression from hybridization or could be the result of homoploid speciation (12). Studies have revealed that ITS is successful for *Piper* species but still, it is yet to prove that ITS2 shows good amplification for *Piper longum* plants. Also, in this study, ITS2 was successfully amplified for the male plant only. Thus, it is early to consider ITS2 to be used as a tool to distinguish between male and female *Piper longum* thereby helping in selecting economically important plants.

BLAST

The sequences obtained by Sanger Sequencing were reverse complemented in

BioEdit software and analyzed using Blastn. BLAST for *rbcL* sequences of the male and female plants revealed maximum similarity to *Piper longum* with 95.07% and 94.66% respectively, and for *psbA-trnH*, both male and female plants showed the maximum similarity to *Piper nigrum* with 95.38% and 95.96% respectively. BLAST using ITS2 showed no similarity with any other species.

Pairwise Sequence alignment-emboss needleman

Pairwise alignment between the male and female *Piper longum* samples showed 89.40% similarity for *rbcL* and 93.4% for *psbA-trnH*. A maximum similarity between the male and female sequences was shown by *psbA-trnH* (Table 1).

Table 1: Pairwise sequence alignment for sequence similarity identification using EMBOSS Needle algorithm for male and female *Piper longum* plants with *rbcL*, ITS2, and *psbA-trnH*

DNA Barcodes/ primers	Length sequences	% of identity	% of similarity	Gap penalty	Extended gap	Gaps	Score
<i>rbcL</i>	624	89.4 (558/624)	89.4 (558/624)	10.0	0.5	30/624	2539.0
ITS2	627	33.8 (212/627)	33.8 (212/627)	10.0	0.5	347/627	324.5
<i>psbA-trnH</i>	331	93.4 (309/331)	93.4 (309/331)	10.0	0.5	11/331	1710.0

Phylogenetic analysis

The UPGMA-based phylogenetic trees were constructed using MEGA-X software for both *rbcL* and *psbA-trnH* sequences. From the phylogenetic tree for *rbcL*, the male *Piper longum* plant was found to be closely related to *Piper nigrum* while the female *Piper longum* was closely related to *Piper attenuatum* by sharing a common ancestor respectively (Figure 4).

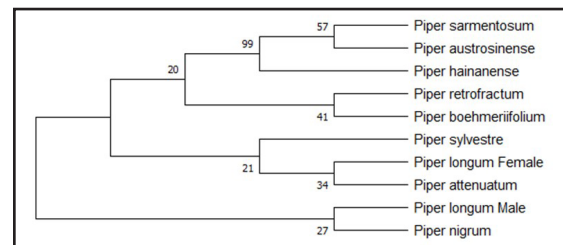


Figure 4: Phylogenetic tree constructed using UPGMA tree method of male and female *Piper longum* plant for *rbcL*

For psbA-trnH, both the male and female *Piper longum* plants have formed a cluster indicating a shared common ancestor which is evident from the highest sequence similarity between male and female *Piper longum* as seen in BLAST analysis (Figure 5).

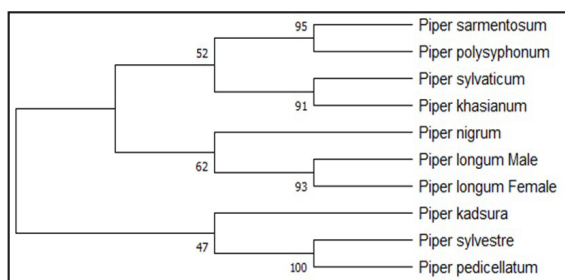


Figure 5: Phylogenetic tree constructed using UPGMA tree method of male and female *Piper longum* plant for psbA-trnH

Conclusion

In this study, the DNA barcoding of *Piper longum* has been successful using rbcL, ITS2, and psbA-trnH. The *Piper longum* samples have shown amplification for all the barcodes used. Among the DNA barcodes, rbcL and psbA-trnH have shown results for both male and female *P. longum* whereas ITS2 has shown significant results for only male *P. longum*. The phylogenetic study has revealed the similarity of *Piper longum* with other *Piper* species. Thus, the DNA barcodes for both male and female *P. longum* were successfully generated for rbcL and psbA-trnH for *Piper longum*.

References

1. Banerjee, N.S., Manoj, P. and Das, M.R. (1999). Male-sex-associated RAPD markers in *Piper longum* L. *Current Science*, 77: 693-695.
2. Viswanathan, T.V. (1995). Medicinal and Aromatic plants. *Advances in Horticulture*, 373-383.
3. Agrawal, V., Sharma, K., Gupta, S., Kumar, R. and Prasad, M. (2007). Identification of sex in *Simmondsia Chinensis* (Jojoba) using RAPD markers. *Plant Biotechnology Reports*, 1(4): 207-210.
4. Kumar, S., Kamboj, J., Suman. and Sharma, S. (2011). Overview of various aspects of the health benefits of *Piper longum* L. fruit. *Journal of acupuncture and meridian studies*, 4(2): 134–140
5. Manoj, P., Banerjee, N.S., and Ravichandran, P. (2005). Development of sex-associated SCAR markers in *Piper longum* L., *PGR Newsletter*, 141: 44-50.
6. Devi, J. and Das, R. (2018). Pipli cultivation: A potential income-generating option, *Journal of Medical Plant Studies*, 6(6): 149-151.
7. Mulcahy, D.L., Weeden, N.F., Kesseli, R. and Carroll, S.B. (1992). DNA probes for the Y-chromosome of *Silene latifolia*, a dioecious angiosperm. *Sex Plant Reproduction*, 5(1): 86-88.
8. Ghariani, S., Trifi-Farah, N., Chakroun, Mohamed. Marghali, S. and Marrakchi, M. (2003). Genetic diversity in Tunisian perennial ryegrass revealed by ISSR markers. *Genetic Resources and Crop Evolution*, 50: 809-815.
9. Hebert, P.D., Cywinska, A., Ball, S.L. and Dewaard, J.R. (2003). Biological identifications through DNA barcodes, *Proceedings. Biological Sciences*, 270(1512): 313-321.
10. Schori, M. and Showalter, A.M. (2011). DNA barcoding as a means for identifying medicinal plants of Pakistan, *Pakistan Journal of Botany*, 43: 1-4.
11. Doyle, J.J. and J. L. Doyle. (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin* 19: 11-15.

12. Lauron-Moreau, A., Pitre, F.E., Argus, G.W., Labrecque, M. and Brouillet, L. (2015). Phylogenetic relationships of American Willows (*Salix L.*, Salicaceae). PLoS ONE, 10(4).